



Normothermic Perfusion in Organ Preservation and Evaluation. Promising Future in Kidney Transplantation.

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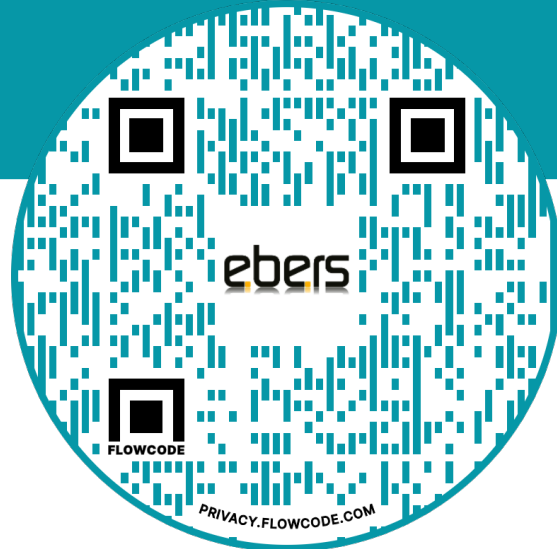
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INTRODUCTION

Kidney transplantation remains the best treatment for end-stage renal disease, but the number of people on the waiting list for a deceased donor kidney is stably high or growing, currently standing at ~10.500 patients in UE in 2024 [1]. To meet growing organ demands, using of grafts donated after circulatory death (DCD), or retrieved from extended criteria donors (ECD), is unavoidable. Unfortunately, DCD and ECD grafts are more immunogenic, which leads to difficulties in immune regulation after implantation and a higher risk of acute and chronic rejection. In response to these needs, the Normothermic Machine Perfusion (NMP) emerges as a tool that resolves these problems. Normothermic Machine Perfusion is an emerging preservation modality for kidneys prior to transplantation that maintains the organ at body temperature (35-37°C), in contrast to traditional cold storage methods. NMP circulates a warm perfusion solution through the renal vasculature, delivering oxygen and nutrients to maintain metabolic activity [2]. This restoration of function during preservation represents a significant advancement in transplantation science, as it allows clinicians to assess organ quality before transplantation and provides a platform for delivering pre-transplant therapies to promote recovery [3].



AR Video To Illustrate
Functionality And
Method Of Use

ADVANTAGE OF NMP OVER TRADITIONAL STORAGE

KEY FEATURE	STATIC COLD STORAGE (SCS)	NORMOTHERMIC MACHINE PERFUSION (NMP)	KEY INSIGHTS AND ADVANTAGES
Principle of Preservation	Reduces metabolic rate to a minimum by hypothermia, halting cellular activity.	Maintains near-physiological conditions (normothermia) with continuous flow of perfusate.	Similar in-vivo environment
Temperature	0-4 °C Hypothermic	34 - 37 °C Normothermic	NMP allows for active metabolic evaluation.
Metabolic State of Organ	Minimal or suppressed	Active and functional (capable of oxygen consumption, ATP production, and waste removal).	Facilitates ex vivo organ reconditioning.
Assessment of Organ Viability	Limited	Dynamic & Real-Time (e.g., O2 consumption). Offers objective viability markers.	Offers objective viability markers and prognosis.
Duration of Preservation	Limited, typically 18-24 hours for kidneys; longer durations increase risk of injury.	Potentially longer preservation times (6 - 48 + hours). Allows for more thorough evaluation.	Reduces ischemia-reperfusion injury.
Therapeutic Interventions	Not possible during preservation.	Allows during preservation: • Drugs administration • Cell therapy (MSC, MAPC) • Gene therapy (siRNA) • Biological therapy (αCD47ab) • Nanotechnologies	Enables organ optimization before transplantation.
Organ Quality/Utilization	May exacerbate injury in marginal organs; limits use of DCD or expanded criteria donor (ECD) kidneys.	Can potentially recondition and improve marginal organs; expands the pool of transplantable organs, especially DCD and ECD.	Increases the number of organs available for transplant.
Complexity/Cost	Low	High (specialized equipment and personnel)	Initial investment often offset by improved clinical outcomes.

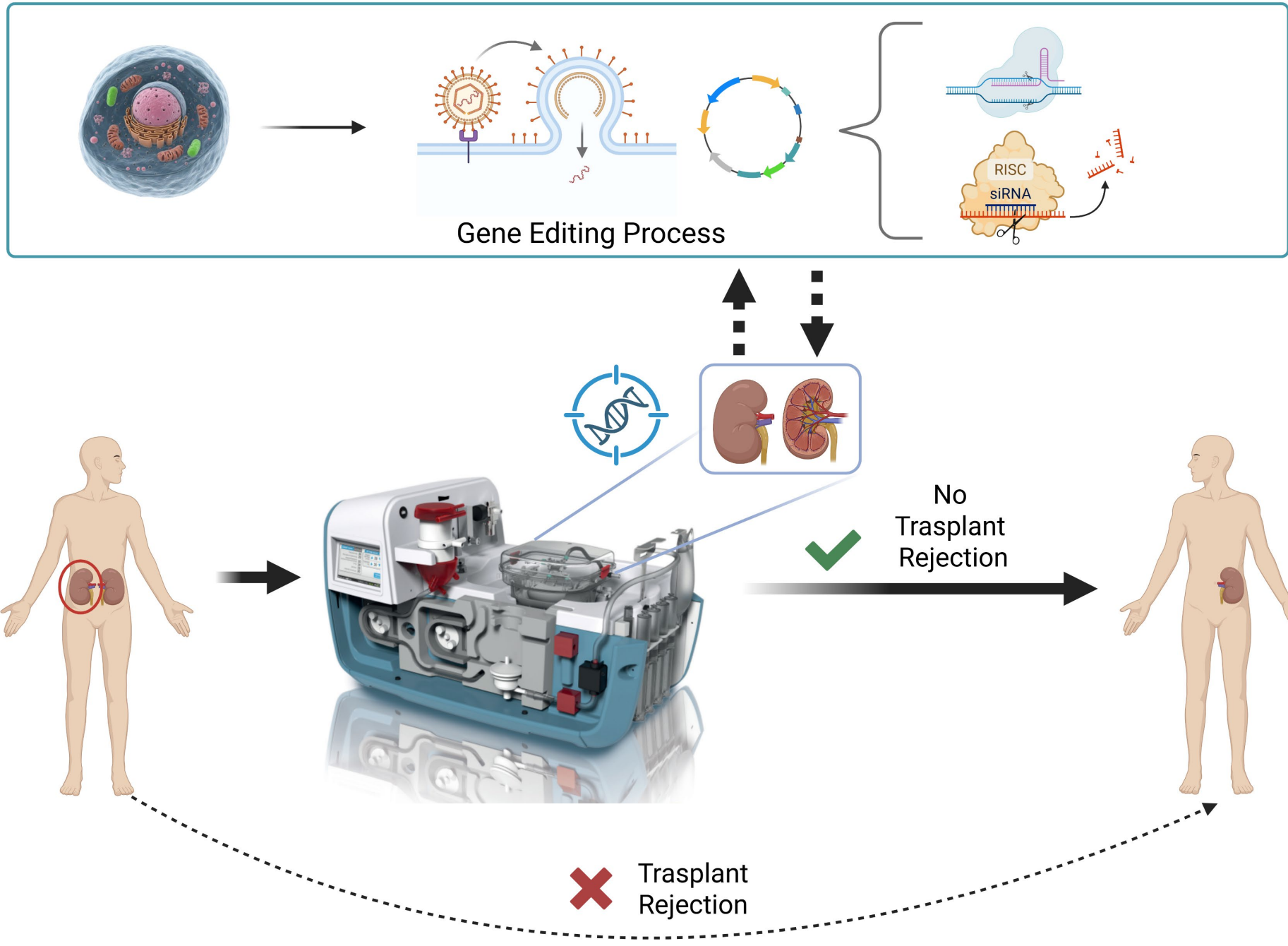
THERAPEUTICS INTERVENTIONS ON NMP AND RESULTS

TARGET MECHANISM/PATHWAY	ORGAN MODEL	MAIN FINDINGS
Mesenchymal Stromal Cell (MSC) [4]	Porcine kidneys	LDH NGAL Regenerative Effect In Ischemia Reperfusion Injury
Anti-CD31 Nanoparticles (NP) [5]	Non-transplantable human kidneys	Anti-human CD31 (endothelium) Abs NPs loaded had enhanced retention
FpGalNAc deacetylase and FpGalactosaminidase through <i>Flavonifractor plautii</i> to enzymatically convert donor blood antigen group A to Group O. [6]	Non-transplantable human kidneys	A antigen loss of aprox. 80% ~ 2h in NMP

OUR FUTURE APPROACH

Our approach, based on the background above context, will be to use the organ *ex vivo* maintenance machine in normothermia **ARK KIDNEY**® of EBERS Medical Technology SL. to perform genetic modifications in kidneys.

With the help of this machine will be able to use gene editing tools such as RNAi or CRISPR-Cas9 using lentiviral vectors, depending on the desired intensity of modification, to try to downregulate the expression of the Major Histocompatibility Complex (MHC) and increase compatibility in kidneys that were not previously compatible.



CONCLUSIONS

- Normothermic machine perfusion is a transformative technology that allows donor kidneys to function and be evaluated outside the body, improving graft viability prior to transplantation.
- This unique capability not only reduces complications such as delayed graft function (DGF), but also makes it possible to use kidneys previously considered marginal or unsuitable.

- NMP offers an innovative platform for research and development of new therapies, allowing for testing the efficacy of treatments or organ repair outside the body prior to transplantation.
- By providing a platform for real-time assessment and therapeutic interventions, NMP promises to expand the donor pool and optimize long-term outcomes for kidney transplant patients.

REFERENCES

[1]. STATISTICS (2025) EUROTRANSPLANT. Available at: https://statistics.eurotransplant.org/index.php?search_type=&search_orga_n=kidney&search_region=All%2BET&search_period=by%2Byear&search_characteristic=&search_text=

[2]. ZAZA, GIANLUIGI et al. "Proteomics reveals specific biological changes induced by the normothermic machine perfusion of donor kidneys with a significant up-regulation of Latexin." Scientific Reports 13 (2023).

[3]. ELLIOTT, TEGWEN R et al. "Normothermic kidney perfusion: An overview of protocols and strategies." American Journal of Transplantation 21 (2020).

[4]. POOL, MEREL B.F. et al. "Treating ischaemically damaged porcine kidneys with human bone marrow and adipose tissue derived mesenchymal stromal cells during ex vivo normothermic machine perfusion." Stem cells and development (2020).

[5]. TIETJEN, GREGORY T. et al. "Nanoparticle targeting to the endothelium during normothermic machine perfusion of human kidneys." Science Translational Medicine 9 (2017).

[6]. MACMILLAN, SERENA et al. "Enzymatic conversion of human blood group A kidneys to universal blood group O." Nature Communications 15 (2024).

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